

# Product Sheet

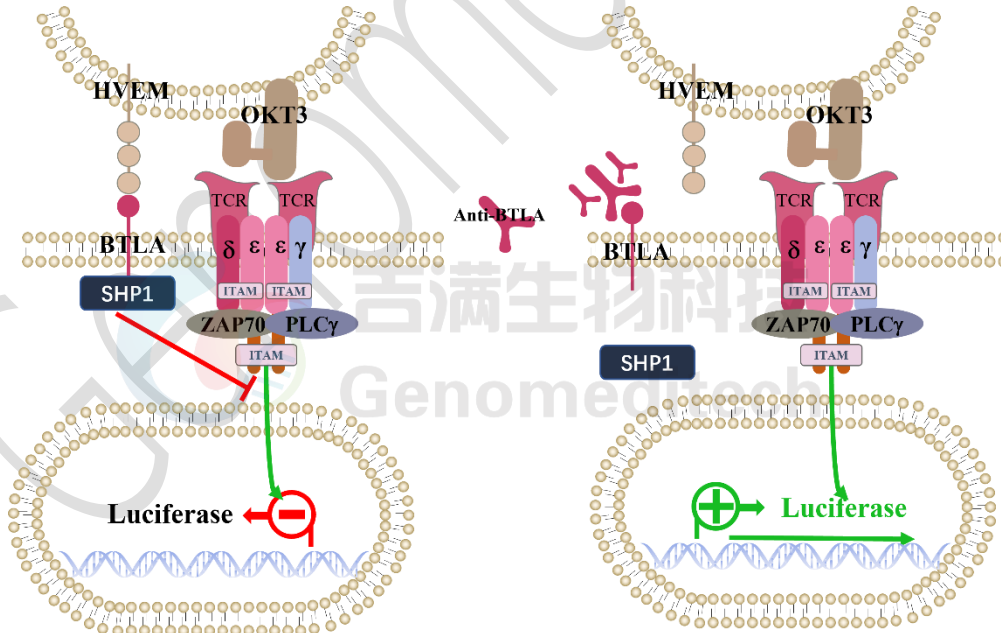
## H\_BTLA Reporter Cell Line

Catalog number: GM-C28122

Version 3.3.1.241205

B and T lymphocyte attenuator (BTLA; CD272) is an Ig superfamily member that negatively regulates immune cell activation, primarily expressed on B cells, T cells, and dendritic cells. Its natural ligand, herpesic virus entry mediator (HVEM; TNFRSF14), is part of the tumor necrosis factor receptor superfamily. BTLA and HVEM modulate T cell and antigen-presenting cell functions through their cell surface expression. BTLA binding to HVEM inhibits T cell proliferation, downregulates the activation marker CD25, and suppresses cytokine production (IFN- $\gamma$ , IL-2, IL-4, IL-10) without causing apoptosis, leading to reduced T cell activation and proliferation.

H\_BTLA Reporter Cell Line is a clonal stable cell line that constitutively expresses BTLA, along with signal-dependent expression of a luciferase reporter gene, developed on a base cell line where HVEM and H\_LIGHT has been knocked out. When H\_BTLA Reporter Cell Line are co-cultured with H\_HVEM aAPC CHO-K1 Cell Line, HVEM binds to BTLA, recruits SHP1, and inhibits TCR signaling, thereby suppressing the expression of luciferase. By adding anti-BTLA antibodies to block the BTLA-HVEM interaction, TCR signaling is restored, allowing for normal expression of luciferase. The luciferase readout represents the activation level of the signaling pathway and can thus be used to evaluate the in vitro effects of drugs related to BTLA.



## Specifications

<b>Quantity</b>	5E6 Cells per vial, 1 mL
<b>Product Format</b>	1 vial of frozen cells
<b>Shipping</b>	Shipped on dry ice
<b>Storage Conditions</b>	Liquid nitrogen immediately upon receipt
<b>Recovery Medium</b>	RPMI 1640+10% FBS+1% P.S
<b>Growth medium</b>	RPMI 1640+10% FBS+1% P.S+3.5 µg/mL Blasticidin+400 µg/mL G418+200 µg/mL Hygromycin+0.75 µg/mL Puromycin
<b>Note</b>	None
<b>Freezing Medium</b>	90% FBS+10% DMSO
<b>Growth properties</b>	Suspension
<b>Growth Conditions</b>	37°C, 5% CO <sub>2</sub>
<b>Mycoplasma Testing</b>	The cell line has been screened to confirm the absence of Mycoplasma species.
<b>Safety considerations</b>	Biosafety Level 2
<b>Note</b>	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

## Materials

Reagent	Manufacturer/Catalogue No.
RPMI 1640	VivaCell/C3010-0500
Fetal Bovine Serum	Cegrogen biotech/A0500-3010
Pen/Strep	Thermo/15140-122
Blasticidin	Genomeditech/ <a href="#">GM-040404</a>
G418	Genomeditech/ <a href="#">GM-040402</a>
Hygromycin	Genomeditech/ <a href="#">GM-040403</a>
Puromycin	Genomeditech/ <a href="#">GM-040401</a>
Clear Flat-Bottom Immuno Nonsterile 96-Well Plates	Thermo/442404
H_HVEM aAPC CHO-K1 Cell Line	Genomeditech/ <a href="#">GM-C25499</a>
Anti-BTLA hIgG4 Antibody(22B3)	Genomeditech/ <a href="#">GM-50103AB</a>
GMOne-Step Luciferase Reporter Gene Assay Kit	Genomeditech/ <a href="#">GM-040503</a>

## Figures

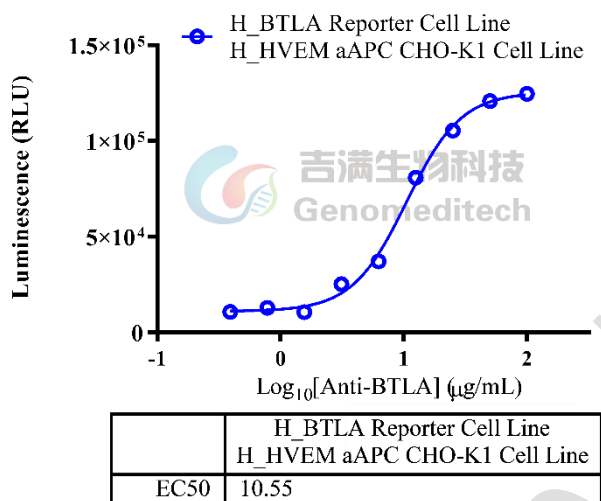


Figure 1 | Response to Anti-BTLA hIgG4 Antibody(22B3). Serial dilutions of the Anti-BTLA hIgG4 Antibody(22B3) (Cat. [GM-50103AB](#)) was incubated with 1E5 cells/well of the H\_BTLA Reporter Cell Line (Cat. [GM-C28122](#)) in a 96-well plate for 1 hour in assay buffer (DMEM + 1% FBS + 1% P.S), then were added to 2E4 cells/well of H\_HVEM aAPC CHO-K1 Cell Line (Cat. [GM-C25499](#)) for 7 hours. Firefly luciferase activity is then measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. [GM-040503](#)). The results indicated a maximum blocking fold of approximately [10.2]. Data are shown by drug mass concentration.

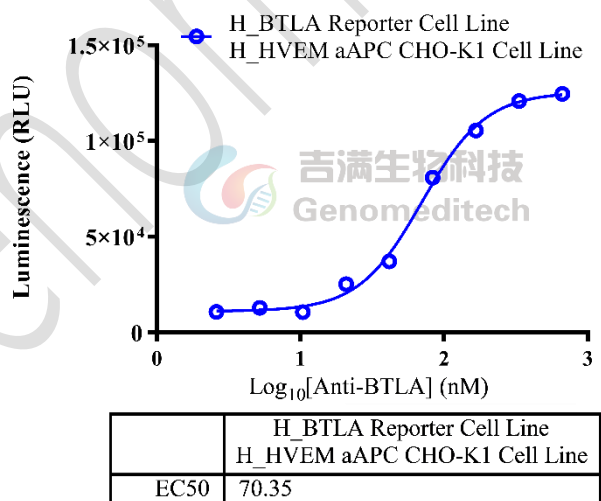


Figure 2 | Response to Anti-BTLA hIgG4 Antibody(22B3). Serial dilutions of the Anti-BTLA hIgG4 Antibody(22B3) (Cat. [GM-50103AB](#)) was incubated with 1E5 cells/well of the H\_BTLA Reporter Cell Line (Cat. [GM-C28122](#)) in a 96-well plate for 1 hour in assay buffer (DMEM + 1% FBS + 1% P.S), then were added to 2E4 cells/well of H\_HVEM aAPC CHO-K1 Cell Line (Cat. [GM-C25499](#)) for 7 hours. Firefly luciferase activity is then measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. [GM-040503](#)). The results indicated a maximum blocking fold of approximately [10.2]. Data are shown by drug molar concentration.

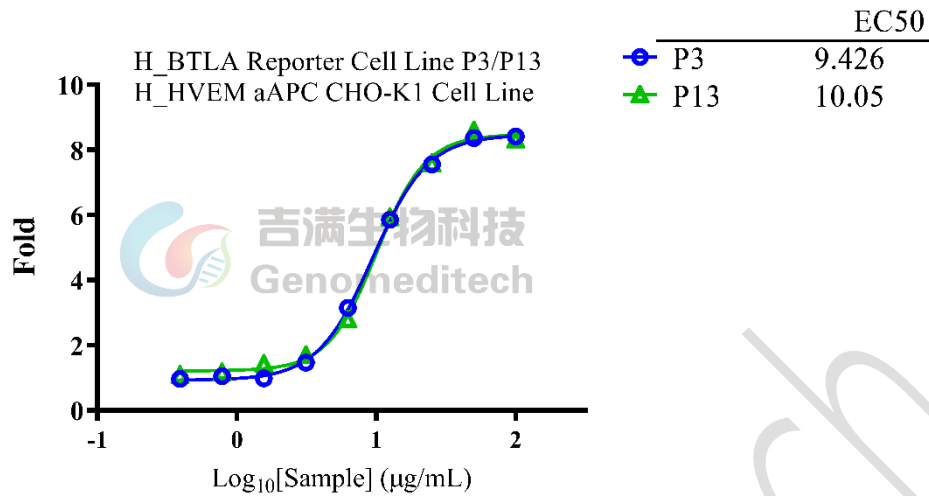


Figure 3 | The passage stability of response to Anti-BTLA hIgG4 Antibody(22B3). Incubate the CP3 and CP13 passage of 1E5 cells/well H\_BTLA Reporter Cell Line (Cat. GM-C28122) with serial dilutions of the Anti-BTLA hIgG4 Antibody (22B3) (Cat. GM-50103AB) for 1 hour, then were added to 2E4 cells/well of H\_HVEM aAPC CHO-K1 Cell Line (Cat. GM-C25499) and incubate in assay buffer (RPMI 1640 + 1% FBS + 1% P.S) for 7 hours. The firefly luciferase activity was measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. GM-040503). Data are shown by drug mass concentration.

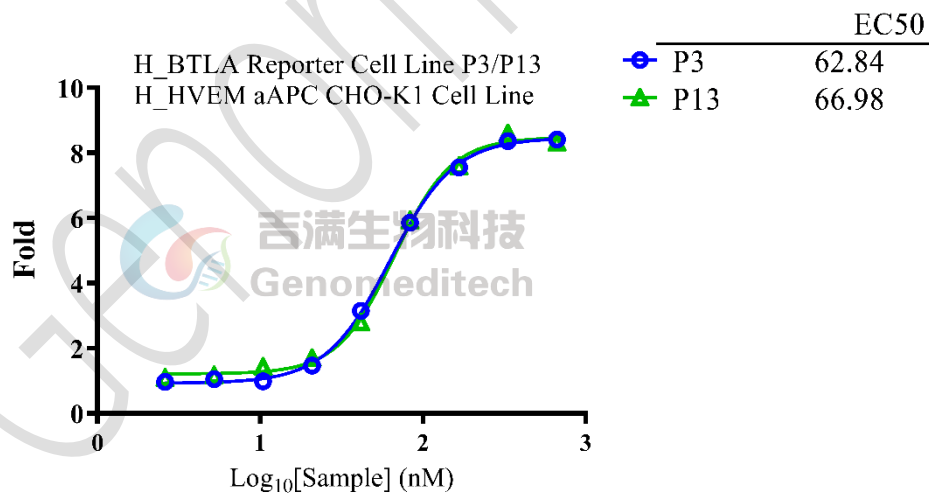


Figure 4 | The passage stability of response to Anti-BTLA hIgG4 Antibody(22B3). Incubate the CP3 and CP13 passage of 1E5 cells/well H\_BTLA Reporter Cell Line (Cat. GM-C28122) with serial dilutions of the Anti-BTLA hIgG4 Antibody (22B3) (Cat. GM-50103AB) for 1 hour, then were added add 2E4 cells/well of H\_HVEM aAPC CHO-K1 Cell Line (Cat. GM-C25499) and incubate in assay buffer (RPMI 1640 + 1% FBS + 1% P.S) for 7 hours. The firefly luciferase activity was measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. GM-040503). Data are shown by drug molar concentration.

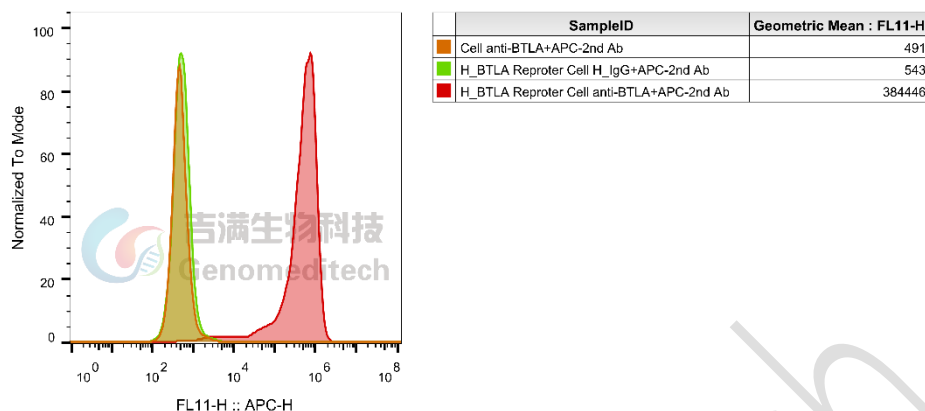


Figure 5 | H\_BTLA Reporter Cell Line (Cat. GM-C28122) was determined by flow cytometry using Anti-BTLA hIgG4 Antibody(22B3) (Cat. GM-50103AB).

## Cell Recovery

Recovery Medium: RPMI 1640+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

- Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).
- Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
- Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium. And spin at approximately 176 x g for 5 minutes. Discard supernatant.
- Resuspend cell pellet with the recommended complete medium. And dispense the suspension into 1 - 2 T-25 culture flasks.
- Incubate the culture at 37°C in a suitable incubator. A 5% CO<sub>2</sub> in air atmosphere is recommended if using the medium described on this product sheet.

## Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

- Centrifuge at 176 x g for 3 minutes to collect cells.
- Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.
- Aliquot 1 mL into each vial.

- d) Place the vial in a controlled-rate freezing container and store at  $-80^{\circ}\text{C}$  for at least 1 day, then transfer to liquid nitrogen as soon as possible.

## Cell passage

Growth medium: RPMI 1640+10% FBS+1% P.S+3.5  $\mu\text{g}/\text{mL}$  Blasticidin+400  $\mu\text{g}/\text{mL}$  G418+200  $\mu\text{g}/\text{mL}$  Hygromycin+0.75  $\mu\text{g}/\text{mL}$  Puromycin

Approximately 48-72 hours after the initial thawing, the cells can be passaged for the first time. After this initial passage, the culture medium can be adjusted to growth medium supplemented with antibiotics. If cells are not passaged within 48 hours, it is recommended to add some fresh recovery medium and place the flask horizontally.

- When the cell density reaches 1.5 - 2E6 cells/mL, subculture the cells. Do not allow the cell density to exceed 2E6 cells/mL.
- It is recommended to use T-25 flasks for subculturing.
- These cells are suspension cells, and it is recommended to use the "half-medium change" method to maintain optimal cell conditions during passaging.
- During passaging, you can directly add fresh growth medium to the culture flask, gently pipette to resuspend the cells, and then transfer the cell suspension to a new T-25 flask for continued culture.

**Subcultivation Ratio: Maintain cultures at a cell concentraion between 3E5 and 1E6 viable cells/mL.**

**Medium Renewal: Every 2 to 3 days**

## Notes

- These cells are sensitive to density, so please ensure that the cell density is maintained within an appropriate range during culture and subculturing.
- During the first passage, pay attention to the nutrient supply; if not subculturing, make sure to add fresh recovery medium every other day as needed.

## Related Products

BTLA:HVEM:LIGHT	
<a href="#">H_BTLA PD-1 Reporter Cell Line</a>	<a href="#">H_HVEM aAPC CHO-K1 Cell Line</a>
<a href="#">H_HVEM PD-L1 aAPC CHO-K1 Cell Line</a>	<a href="#">H_HVEM Reporter Jurkat Cell Line</a>
<a href="#">Cynomolgus_BTLA HEK-293 Cell Line</a>	<a href="#">H_BTLA CHO-K1 Cell Line</a>
<a href="#">H_BTLA HEK-293 Cell Line</a>	<a href="#">H_LIGHT(TNFSF14) CHO-K1 Cell Line</a>
<a href="#">H_TNFRSF14(HVEM) CHO-K1 Cell Line</a>	
<a href="#">Anti-BTLA hIgG4 Antibody(22B3)</a>	<a href="#">Anti-BTLA hIgG4 Antibody(Icatolimab)</a>
<a href="#">Anti-TNFRSF14(HVEM) hIgG4 Antibody</a>	<a href="#">Anti-TNFSF14 hIgG4 Antibody</a>

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